

Product Manual Cat. No: #2001

Taq 2x PCR Master Mix

Description

The Taq 2x PCR Master Mix contains all the components necessary for PCR, including a Taq DNA polymerase variant and an optimized buffer including ultrapure dNTPs.

Taq 2x PCR Master Mix is a ready to use reaction mix. It contains all components necessary for a successful and reliable PCR or primer extension reaction in all standard PCR cyclers. Only primers and template need to be added.

This mix provides robust PCR performance for a wide range of PCR applications. The pre-ready 2x mix ensures reproducible results, significantly reduces setup times and the risk of pipetting mistakes.

Applications include standard PCR, real-time-PCR (addition of suitable dye or probe required), primer extension reactions, TA cloning, 3'A-tailing of blunt ends, and screening / high-throughput PCRs.

Kit components

Component	M pack*	
Taq 2x PCR Master Mix	1x 1.25 mL	

^{*}Other pack sizes, bulk orders and customization are available upon request.

Storage and shipment

Transport with cool packs. The reagents should be stored at -20°C upon arrival. The reagents are stable until the expiration date if stored correctly.

Reaction Master Mix set-up

The recommended master mix set-up for a 50 μ L reaction volume is shown in the table below.

Reagent	Volume (μL)	Final concentration	
Taq 2x PCR Master Mix	25	1x	
∞Forward primer (10 µM)	1	0.2 μM (0.05–1 μM)	
∞Reverse primer (10 µM)	1	0.2 μM (0.05–1 μM)	
Template / Sample extract	x	<1000 ng* DNA	
Nuclease-free water	Up to 50 μL final volume		

Keep all components on ice.

Spin down and mix all solutions carefully before use.

∞Primers should ideally have a GC content of 40–60% typically.

Instrument and program set-up

Cycles	Steps	Temperature	Time
1	Initial denaturation	95°C	2 min
25–40	Denaturation	95°C	15 sec
	Annealing*	54-72°C	30 sec
	Extension	72°C	1 min /1000 bp

^{*}Typically, the annealing temperature is about 3–5°C below the calculated melting temperature of the primers used.

^{*}Suggested template concentration should be about 1 ng – 1000 ng (genomic DNA) or 1 pg – 1 ng (plasmid/viral DNA) per reaction.



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Technical information and support

For technical enquiries or assay development support, please contact us via e-mail at: mdx@medixbiochemica.com.

Additional information and technical resources are available on our website at: info.medixbiochemica.com/resources.